

Cartilagineol, the Fourth Lineage of Laurencia-derived Polyhalogenated Chamigrene

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Abstract: A revised regiochemistry and the absolute configuration are established for cartilagineol (formerly allo-isoobtusol). This revised structure renders cartilagineol the first example of the fourth stereochemical type of Laurencia-derived polyhalogenated chamigrene in support of current biogenetic theory. Earlier misassigned NMR data for ma'ilione is corrected. © 1998 Elsevier Science Ltd. All rights reserved.

A recent paper by Juagdan et al. 1 described two new chamigrene metabolites, allo-isoobtusol (1) and ma'ilione (2), isolated as minor components of a Hawaiian collection of Laurencia cartilaginea. Guella et al. 2 have questioned the structural assignment for allo-isoobtusol suggesting structure 3 as a better fit to the physical data as well as to biogenetic theory. We have isolated both of these compounds, along with known vinyl bromides 4 and 5,3-5 from Laurencia sp. collected at Taytay, Philippines. We wish to report that 3 is the correct structure for allo-isoobtusol, rechristened cartilagineol. 2

Cartilagineol (15 mg), colorless crystals, mp 62-63°, $[\alpha]_D$ -32° (c 0.253, CHCl₃), was separated from 4 by PTLC on silica (hexane:isopropanol, 45:1). Its molecular formula was established as $C_{15}H_{23}Br_2ClO$ by HREIMS. The ¹H and ¹³C NMR spectral data (Table 1) were identical to those reported, ¹ but the assignments for the halogenated carbons C-8 and C-9 have been reversed. Based on carbon chemical shift values and the HMQC spectrum, the quaternary halide at C-9 can be assigned as chlorine (δ_C 73.3) and the methine halide at C-8 as bromine (δ_C 57.1). Although bromine and chlorine display similar α -shift effects at C-1 in 1-methyl-1,2-dihalocyclohexanes, the α -shift effects at C-2 are significantly different; the bromomethine carbon generally appears at least 5 ppm upfield of its chloro analog.⁶

To provide further evidence for both the regio- and stereochemistry of cartilagineol, an x-ray crystal structure was carried out. Crystals of 1 were grown by slow evaporation of a hexane solution at 5°C. A crystal 0.4 x 0.2 x 0.15 mm was selected and mounted on a glass fiber for data collection. The crystal decomposed fairly quickly in the x-ray beam (38% decrease in standard reflection intensity after 25 hrs) and the data collection was terminated early. The compound crystallized in the monoclinic space group P2₁ (a=15.963Å, b=7.118Å, c=16.427Å, β=90.58°) with two independent molecules in the asymmetric unit (Z=4). Only the Br, Cl and O atoms were refined anisotropically. The final least squares refinement gave R=0.126 for 1534 F>4σ (0.152 for all 2097 data) and 194 parameters. While the data set was insufficient for complete refinement, the relative stereochemistry of the molecule is clear and shown in Fig. 1. The hydrogens at the stereogenic centers (in calculated positions) have been included for clarity. Both molecules in the asymmetric unit show the same stereochemistry.

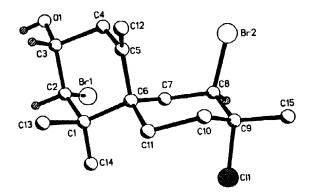


Figure 1: Molecular Structure of 3

Although the x-ray data collection did not allow the determination of the absolute configuration of cartilagineol, several other lines of evidence strongly support the configuration shown in 3 (2S, 3R, 6S, 8R, 9R). First, as pointed out by Guella et al., 2 the optical rotation of cartilagineol is essentially the same as that of isoobtusol (6) (+33°) but opposite in sign. 8 As the interchange of halogens at C-8 and C-9 does not affect the rotation, 2.9 this indicates that 3 and 6 have an enantiomeric relationship. Second, the optical rotation of 5 isolated from the present extract was -38° in agreement with the literature value of -33°.4.10 The absolute configuration of 5 follows from its spontaneous equilibration with 4, whose absolute configuration has been established by x-ray. 3b Third, when we treated cartilagineol (3) with Zn/AcOH, (-)-7 (deschloroelatol) was produced. Both enantiomers of 7 are known8.11 and their absolute configurations are established by their method of preparation: the (-)-isomer from LiAlH4 reduction of obtusol (8) and the (+)-isomer from Zn/AcOH reduction of isoobtusol (6).8

That cartilagineol is correctly represented by structure 3 is of biogenetic significance. Current theory predicts the existence of four stereochemical types for the polyhalogenated *Laurencia*-derived chamigrenes;^{2,12} to date, three types have been reported. Examples of these three types are obtusol (8), isoobtusol (6), and rogiolol (9), all found in distinct *Laurencia* collections. Cartilagineol represents the missing fourth stereochemical type. Its discovery lends credence to the biogenetic theory and to the possibility that *Laurencia*'s secondary metabolites may be useful in sorting out the confused taxonomy of this genus.²

In addition to cartilagineol, ma'ilione (2) (20 mg) and vinyl bromides 4 (4 mg) and 5 (11 mg) were isolated from the extract. Ma'ilione, colorless crystals, mp 135-136°, $[\alpha]_D$ -20° (c 0.196, CHCl₃), was obtained by PTLC on silica (hexane:isopropanol, 15:1). Its NMR spectral data were identical to those reported. However, the original data contained several misassignments which are corrected in Table 1. In particular, the enone vinyl hydrogens were reversed as were the C-13 methyl and the C-11 methylene carbons. The HMQC spectrum clearly distinguishes these latter two carbons. Our optical rotation value was also significantly different from that reported (-20° vs. -100°) and may possibly be explained by the larger size of our sample (9.8 mg vs. 0.4 mg) and its greater purity (crystals vs. oil).

Enone 10, $[\alpha]_D + 9.4^\circ$ (c 0.05, CHCl₃), appears as an artifact produced during the purification of cartilagineol. HRFABMS established its formula as $C_{15}H_{22}BrClO$. Its structure was deduced from an analysis of the ¹H, ¹³C and COSY spectra. ¹³ In the ¹H NMR spectrum of 10, the C-2 and C-3 methine signals and the terminal vinyl hydrogen signals of cartilagineol were replaced by a deshielded vinyl hydrogen (δ 5.84) and a vinyl methyl (δ 2.19). HBr loss followed by isomerization would convert 3 to 10, and all of the spectral data support this conclusion. Such a process has ample precedence in these systems.^{3a,9,14,15}

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Table 1: 13C and 1H NMR Data for Cartilagineol 3 and Ma'ilione 2 in CDCl₃

3			2	
Pos.	13Ca	1 H b	13Ca	1 H b
1	43.7 (s)		42.7 (s)	_
2	76.4 (d)	4.38 (dd, 1.7, 3.7)	68.8 (d)	4.58 (d, 3.0)
3	69.6 (d)	3.65 (br dddd, 3.7, 4.1, 11.3, 12.1)	71.7 (d)	4.18 (m)
4	39.4 (t)	2.42 (br dd, 4.0, 12.1)	38.4 (t)	2.68 (dd, 3.5, 13.0)
		2.70 (t, 12.1, 12.1)		2.71 (br d, 13.0)
5	142.2 (s)		141.7 (s)	_
6	44.0 (s)	_	51.8 (s)	- ·
7	33.7 (t)	2.79 (dd, 4.1, 16.0)	152.9 (d)	7.00 (dd, 1.4, 10.8)
		3.06 (br d, 15.9)		
8	57.1 (d)	4.44 (br s)	131.1 (d)	6.10 (d, 10.6)
9	73.3 (s)	-	198.7 (s)	-
10	32.4 (t)	1.77 (ddd, 4.5, 5.5, 12.7)	34.2 (t)	2.34 (2H, m)
		2.32 (ddd, 4.8, 12.7, 13.0)		
11	24.3 (t)	1.70 (ddd, 4.5, 6.1, 13.7)	26.3 (t)	2.15 (m)
		2.02 (dt, 4.8, 13.5, 13.5)		2.20 (m)
12	114.7 (t)	4.95 (s)	117.9 (t)	4.84 (s)
		5.19 (s)		5.17 (br s)
13	23.3 (q)	1.03 (s)	26.9 (q)	1.04 (s)
14	24.4 (q)	1.29 (s)	21.4 (q)	1.29 (s)
15	33.1 (q)	1.74 (s)		
ОН		2.23 (d, 11.2)		2.26 (br s)

^a Recorded at 50 MHz, CDCl₃ as internal standard (δ 77.0). Multiplicity was based on the JMODXH spectra.

REFERENCES AND NOTES

- 1. Juagdan, E.G.; Kalidindi, R.; Scheuer, P. Tetrahedron 1997, 53, 521-528.
- 2. Guella, B.; Öztunç, A.; Mancini, I; Pietra, F. Tetrahedron Lett. 1997, 38, 8261-8264.
- 3. a) Suzuki, M.; Kurosawa, E. Tetrahedron Lett. 1978, 4805-4808; b) Suzuki, M.; Furusaki, N.; Hashiba, N.; Kurosawa, E. Tetrahedron Lett. 1979, 879-882.
- 4. Schmitz, F.J.; Michaud, D.P.; Schmidt, P.G. J. Am. Chem. Soc. 1982, 104, 6415-6423.
- 5. Rashid, M.A.; Gustafson, K.R.; Cardellina II, J.H.; Boyd, M.R. Nat. Prod. Lett. 1995, 6, 255-259.
- 6. Crews, P.; Naylor, S.; Hanke, F.J.; Hogue, E.R.; Kho, E.; Braslau, R. J Org. Chem. 1984, 49, 1371-1377.
- 7. a) Foxman, B. M. Inorg. Chem. 1978, 17, 1932-1938; b) Foxman, B. M.; Mazurek, H. Inorg. Chem. 1979, 18, 113-116.
- 8. a) González, A.G.; Darias, J.; Díaz, A.; Fourneron, J.D.; Martín, J.D.; Pérez, C. Tetrahedron Lett. 1976, 3051-3054; b) González, A.G.; Martín, J.D.; Martín, V.S.; Martínez-Ripoll, M.; Fayos, J. Tetrahedron Lett. 1979, 2717-2718.
- 9. Guella, G.; Mancini, I.; Chiasera, G.; Pietra, F. Helv. Chim. Acta 1990, 73, 1612-1620.
- 10. A compound reported as the enantiomer of 5 was drawn as its diasteromer (Coll, J.C.; Wright, A.D. Aust. J. Chem. 1989, 42, 1591-1603). Because its optical rotation was -40°, it is likely this compound is 5.
- 11. Kennedy, D.J.; Selby, I.A.; Thomson, R.H. *Phytochemistry* **1988**, 27, 1761-1766. See König, G.M.; Wright, A.D. *J. Nat. Prod.* **1997**, 60, 967-970 for reassignments of ¹H NMR values.
- 12. Sakai, R.; Higa, T.; Jefford, C.W.; Bernardinelli, G. Helv. Chim. Acta 1986, 69, 91-105.
- 13. IH NMR (CDCl₃, 200 MHz) δ 1.02 (s), 1.15 (s), 1.79 (s), 2.07 (dd, 4.5, 15.4 Hz), 2.12 (br d, 18.3 Hz), 2.19 (d, 1.2), 2.22 (m), 2.28 (m), 2.33 (m), 2.38 (m), 2.47 (br d, 18.2), 2.92 (dd, 4.7, 15.6), 4.62 (dd, 4.5, 7.6), 5.84 (br s); ¹³C NMR (CDCl₃, 50 MHz, carbonyl carbon not observed) δ 23.9 (t), 24.1 (q), 26.1 (q), 28.5 (q), 28.8 (q), 37.0 (t), 39.4 (t), 41.9 (s), 44.6 (s) 49.2 (t), 58.5 (d), 71.3 (s), 127.9 (d), 168.6 (s).
- 14. Brennan, M.R.; Erickson, K.L.; Minott, D.A.; Pascoe, K.O. Phytochemistry 1987, 26, 1053-1057.
- 15. González, A.G.; Martín, V.S.; Norte, M. Tetrahedron Lett. 1979, 2719-2722.

b Recorded at 500 MHz.